Comparative Assessment of Growth Performance and Nutrients Utilization of African Catfish (*Clarias gariepinus*, Burchell 1882) Fed Chicken Offal and Shrimp-Based Diets

Elvis Monfung Ayim¹, Ettah Akpang Ivon², Raymond Odey Ajang¹ and Akaninyene Paul Joseph¹*

¹Department of Zoology and Environmental Biology, Faculty of Biological Sciences, University of Calabar, Calabar, Nigeria.

²Department of Science Laboratory Techniques, Faculty of Biological Sciences, University of Calabar, Calabar, Nigeria.

Authors’ contributions

This work was carried out in collaboration among all authors. Author APJ designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Authors EMA and EAI managed the analyses of the study. Author ROA managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/ARRB/2018/v30i530026

Original Research Article

ABSTRACT

A comparative study on the growth performance of African catfish (*Clarias gariepinus*) fed shrimp based diet (SBD), chicken offal based diet (COBD) and coppens feed was carried-out for 22 weeks in concrete tanks measuring 3.5 x 1.7 x 1.5 m³. Triplicate groups of 40 juveniles with average length of 9.15 ± 0.17cm and weight of 20.00 ± 2.58g were stored in each tank. A total of 360 juveniles were used throughout the study. Fish were fed daily at 3% of their body weight through-out the duration of the experiment. The nutrient composition of the 3 nutrients differed significantly at p<0.05. Weight gain (WG) (g) was highest (17189.33 ± 506.61g) in fish fed diet A (Coppens feed), followed by 15045.33±202.42 g (diet B) (SBD) and least (14357.33 ± 108.25 g) (diet C) (COBD). The Growth

*Corresponding author: E-mail: joseph.akan@yahoo.com;
rate (GR) was highest in fish fed with diet A (111.62 ± 3.29 g/day), followed by 97.69±1.31 g/day (diet B), and least (93.23±0.70 g/day) in diet C. Specific growth rate (SGR) was highest in juveniles fed diet A (2.02 ± 0.02 %/day), followed by 1.94±0.01 %/day for diet B, and least (1.91±0.01 %/day) for diet C. Mean growth rate (MGR) was highest in juveniles fed diet A (11.88 ± 0.29 mg/day), followed by 11.74±0.15 mg/day for diet B, and least (11.69±0.09 mg/day) in fish fed diet C. The WG, GR and SGR varied significantly between the juvenile fish group fed the 3 diets at p<0.05, while MGR did not vary significantly between fish group fed the 3 diets at p>0.05. Fish fed diet A had a higher feed consumption (FC) (41650.00 ± 315.34 g/kg), than that fed diet B (39034.24 ± 86.34 g/kg) and then diet C (38276.00 ± 342.97 g/kg). Juveniles fed Coppens feed did better, with a feed conversion ratio (FCR) of 2.42 ± 0.06, followed by 2.59±0.04 for diet B and 2.67 ± 0.03 for fish fed diet C. Feed conversion efficiency (FCE) was higher in fish fed diet A (41.26±0.94 %), followed by 38.54 ± 0.54 % (diet B) and least (37.52 ± 0.51 %) (diet C). The FC and FCR varied significantly between the fish fed the 3 diets at p<0.05, while FCE was not significantly different between the 3 treatment groups at p>0.05. The water parameters of the culture water varied significantly between each treatment groups (p<0.05), and were within the rage suitable for normal growth of fish. Though coppens feed yielded better growth performance and feed utilisation, the use of COBD and SBD is recommended for fish farmers in Nigeria. More researches should be carried out on using chicken offals and shrimps in fish feed formulation.

Keywords: Growth performance; feed utilization; feed composition; Clarias gariepinus; coppens; chickel offal and shrimp based diet.

1. INTRODUCTION

Clarias gariepinus has been widely regarded as one of the most relevant tropical fresh water species of fish for aquaculture, because of the potentials they exhibit as documented by [1]. Nutritionally, this fish is rich in vitamin D, low in Omega-3 fatty acids and high in Omega-6 fatty acids, which promote growth and brain development. It has a fast growth performance rate based on tolerance of adverse ecological condition. The primary objectives of the aquaculture industry are to optimize growth and make available high-quality fish [2], and with an increase in the World-wide awareness of fish as a treasured source of protein, has led to an increase in the prospects of aqua feeds with diets being specially design to meet the nutritional needs of species, life cycle and the health conditions of fish [3]. Fishmeal has been the most used source of protein incorporated in culture fish diets, despite the fact that, fish supply is not increasing globally [4]. The prerequisite for the development of large-scale aquaculture is discovering locally formulated feeds of high-quality protein to replace fish meal. Many fish nutritionist have studied alternative sources of fish protein, both from plant and animal with some successes [5,6,7,8], some of which include earthworm and termite (Macrotermes Subhyalinus) meals [9], Maggot meal [10,11] and Phane (Imbrasia belina), insect [12] to replace fishmeal that is desired as the chief source of protein in diets of fish, primarily due to its high digestibility and rich essential amino acids (EAAS). Essential amino acids cannot be synthesized in required quantities necessary for fish growth and tissue development, hence, must be fortified in diets of fish [13]. Re-cycled chicken meal is termed as the dry rendered product from an aggregate of chicken flesh, skin and bone obtained from whole carcass of chicken with the exception of feathers, which are ground and reduced in particle size [14]. Aquaculture possesses the capacity to become a sustained farming practice which can add to captured fisheries, and considerably help in feeding the growing population of the World [15]. In the year 2002, about 70% of global production of reared fish was recorded to have been in China alone [16]. Udo et al. [17] concurred that for the ingredients to be introduced into least-cost ration, there is need for the evaluation of nutrients, as it exposes several key aspects for different experimental methods used. Many researchers [18] and [19] stated that knowledge of the ingredients to be used in the formulation of feed for the African catfish is essential, particularly about ingredient digestibility, specification, palatability. Many researchers [20] and [21] purported that, the nutritional constituents of chicken offal meal can vary depending on the part being processed. Fish feed remain one of the major input in fish production. In Nigeria, the major challenge confronting the development and growth of aquaculture is lack of feed, the technology of fish feed production is little developed in Africa and some developing
Countries [22]. The study was aimed at assessing the growth performance of African Catfish (C. gariepinus) juveniles fed with chicken offal and shrimp-based diets.

2. MATERIALS AND METHODS

2.1 Study Area

This research took place at Andem and Sons Fish Farm Limited, located in Calabar South Local Government Area, Calabar, Cross River State, Nigeria.

2.2 Experimental Design

This research work was undertaken for 22 weeks (i.e. five months and two weeks); between June-November, 2016. Nine concrete tanks measuring (3.5 x 1.7 x 1.5m$^3$) were used. The 9 concrete tanks were used, labelled A$_1$, A$_2$, A$_3$, B$_1$, B$_2$, B$_3$ and C$_1$, C$_2$, C$_3$, making 9 experimental units to aid replication of the experiment. Two different types of experimental fish diet were formulated with the addition of Chicken offal (diet C), shrimp waste (diet B), which were used to compare with Coppens commercial feed (control) (diet A). This scientific study was conducted in replicates of 3, using fish juveniles with the average length of 9.15±0.17cm and average weight of 20.00±2.58g. The experimental fishes were fed twice a day between 7:00 and 8:00am and 6:00 and 7:00pm at 3% of their body weight.

2.3 Collection, Preservation and Preparation of Chicken Offals and Shrimp Waste

The chicken offal used in this study was purchased from the Watt Market and Mr. Runyi broiler slaughtering farm at the Cross River State Water Board premises, all in Calabar, Cross River State.

The freshly collected offal was thoroughly washed in water carefully to remove the faecal content as much as possible before weighing. The Chicken offal was then par boiled for 30 minutes. It was allowed to cool and then sun dried. Shrimp waste was obtained at the Calabar beach market in the dry form, it was then packed in a sack bag and kept in a dry place until when needed.

2.4 Diets Ingredient and Diet Formulation Methods

Experimental diet were composed of Soyabean meal (SBM), Chicken offal (CO), Shrimp meal (SHM), Wheat offal (WO), Cassava starch, vitamin premix, bone ash/calcium, Sodium chloride (NaCl), vegetable oil, lysine, methionine.

Two diets were formulated for this experiment, adopting Pearson square system to arrive at a crude level of protein of 42%. Various ingredients of feed were grinded and amalgamated properly in accordance with their percentages. After which the feed were pelletised using a Hand Cranker machine into small sizes and oven dried. Soon after drying was over, the feed were packed in bags and stored. The processes of the feed formulation were all carried out at Aqua Marvels Farms in Calabar. The farm is also a designated centre (Nigeria Markets II) used by the United State Agency for International Development (USAID) for Wet Field Demonstration on improved Aquacultural Practices, which was established in 2014. The components and proportions of ingredients making-up the diets were:

<table>
<thead>
<tr>
<th>Ingredients</th>
<th>Amount in g (%)</th>
<th>Amount in g (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chicken offal (CO)</td>
<td>---</td>
<td>370</td>
</tr>
<tr>
<td>Shrimps meal (SHM)</td>
<td>360 36</td>
<td>--- 37</td>
</tr>
<tr>
<td>Soybeans meal (SBM)</td>
<td>360 36</td>
<td>370 37</td>
</tr>
<tr>
<td>Yellow maize (YM)</td>
<td>120 12</td>
<td>110 11</td>
</tr>
<tr>
<td>Wheat offal (WO)</td>
<td>120 12</td>
<td>110 11</td>
</tr>
<tr>
<td>Methionine</td>
<td>2.5 0.25</td>
<td>2.5 0.25</td>
</tr>
<tr>
<td>Lysine</td>
<td>2.5 0.25</td>
<td>2.5 0.25</td>
</tr>
<tr>
<td>Bone ash/calcium</td>
<td>5 0.5</td>
<td>5 0.5</td>
</tr>
<tr>
<td>Vitamin premix</td>
<td>15 1.25</td>
<td>15 1.25</td>
</tr>
<tr>
<td>Sodium chloride (NaCl)</td>
<td>5 0.5</td>
<td>5 0.5</td>
</tr>
<tr>
<td>Cassava Starch</td>
<td>5 0.5</td>
<td>5 0.5</td>
</tr>
<tr>
<td>Palm oil</td>
<td>5 0.5</td>
<td>5 0.5</td>
</tr>
<tr>
<td>Total in g/kg</td>
<td>1kg</td>
<td>1kg</td>
</tr>
</tbody>
</table>

Diet B = Shrimp-based diet (SBD), Diet C = Chicken Offal-based diet (COBD)
2.5 Coppens Feed Composition and Characteristics

Coppens feed produced by Coppens International in Netherlands is made up of good standard ingredients like calcium, methionine, copper sulphate (CuSO4), marine fish meal, phosphorus, lysine, selenium refined fish oil and several grains. Sizes of coppens feed used were 3mm, 4mm and 6mm respectively.

2.6 Tank Treatment

Prior to the commencement of the experiment, the tanks were treated with common salt sodium chloride (NaCl) for complete extermination of micro-organisms which can pose a threat to the juveniles. After which water was filled and allowed for two weeks, then flushed out, before the refilling and introduction of the juveniles.

2.7 Fish stocking and Experimental Procedures

Borehole water (ground water) was the source of water, was pumped into the tanks by electrical pump and piped into the experimental tanks. Three hundred and sixty (360) juveniles were obtained from the University of Calabar Fish Farm Hatchery complex, Institute of Oceanography, Calabar, and transported to Andem and Sons fish farm in 50 liter water storage tank, where they were allowed to acclimate to the new environmental conditions for about 2 weeks. During this period, they were fed twice a day between 7:00 and 8:00am and 6:00 and 7:00pm at 3% of their body weight. Forty fish juveniles with average length of 9.15±0.17cm and average weight of 20.00±2.58g were stocked per unit. Before stocking, the initial length and weight of each fish sample was accurately measured using a measuring board (to the nearest centimeters) and electronic weighing balance (Metlar mt-5000D version) (to the nearest grams) respectively. The culture water was changed every 48 hours, in order to maintain good water quality through-out the experiment.

The fish were sampled bi-weekly to determine their growth and survival (mean body weight, mean total length and mortality). The rations was always adjusted so as to correspond with the new body weight using the weighing balance. The sampling exercises were carried out between 7-8am to minimize heat stress [23].

2.8 Monitoring of Water Quality Parameters

Water quality parameters like; dissolved oxygen (DO), hydrogen ion concentration (pH), temperature (°C), ammonia (NH3) were measured bi-weekly, through-out the experiment. Mercury-in-glass thermometer was used for measuring water temperature in degrees celsius. The pH and oxygen concentrations were measured in-situ using the Jenway meters; model 3050, England for DO in milligram per liter (mg/L) and model 9070 for pH. Water was collected into a 1 liter container, preserved in an ice chest before taking to the physico-chemical laboratory of the Cross River State Water board, Calabar for ammonia level determination using spectrophotometer in mg/L.

2.9 Proximate examination of the Experimental Diets

The proximate analysis of the formulated diets were determined according to standard methodology of [24]. Crude protein content was determined using Kjeldahi method. All proximate analysis were carried-out in the Faculty of Agriculture Central Laboratory, University of Calabar.

2.9.1 Determination of moisture level

A neat crucible was subjected to drying in an oven to a constant weight (a) before introducing a quantity of the sample into a beaker, then weighed (b). Next, the sample was dried inside a ventilated heated oven which was powered electrically at 75°C for 24 hours, then allowed to cool in a desiccator, and then weighed. The procedure was repeated until a constant weight (c) was reached. Same procedure was carried out 3 times for each sample. The percentage moisture level was mathematically calculated using the formula:

\[
\% \text{ moisture content} = \frac{b-c}{b-a} \times 100\%
\]

2.9.2 Ash content

The crucible was ignited at 550°C for 3 hours, then cooled and weighed. Five grams of the sample was placed in the crucible and weighed. It was then burnt at 550°C for a day, cooled and then weighed. Same procedure was carried out over and over again until a constant weight was obtained. The calculation of
percentage ash content followed the formula as shown below:

\[
\% \text{ Ash content} = \frac{\text{wt of ash}}{\text{wt of sample}} \times 100
\]

2.9.3 Crude fat or ether extract

Five grams of the sample was weighed and put in a thimble. About 120 mL of petroleum ether was emptied into an earlier dried and weighed round bottom flask. The thimbles and its content were introduced into an extractor known as the soxhlet extractor, which became fitted into the spherical bottom flask and then condensed together with the extraction apparatus to allow the sitting of the flask on the spaces provided in the hot plate. The hot plate was set to gentle heat. With tap on, the ether evaporated and condensed, dropping into a thimble from where it extracted the soluble ether contents into a round bottom flask. The process continued for 10 hours, the thimble was removed and dried in the air, later the fat from the extract was utilized for the determination of fibre. Then, petroleum ether present in the flask was distilled off and received in the soxhlet extractor tube. Drying of the flask was carried out in an air circulating desiccator for two days. The circular bottom flask with the lipid extract inside was then weighed. The content inside the flask was dried and weighed to a constant weight. The lipid quantity that was obtained from the difference between the flasks weighed previously and later-on was obtained as shown below:

\[
\% \text{ Ether Extract} = \frac{\text{wt of extract}}{\text{wt of sample}} \times 100
\]

2.9.4 Crude fibre

For acid digestion, the fat free material (8-10g) was weighed and transferred into a 400mL beaker that was previously marked at 200 mL level. Fifty milliliters of sulphuric acid (i.e. 1.25%) was added and the mixture rose to 200 mL marked. The beaker together with the content was heated to a boiling point for half an hour. The content of the beaker was then filtered through a Buchner funnel with the aid of a suction pump. The residue was washed with hot water until it was acid free. For base digestion, the residue left after acid digestion was transferred into 400 mL beaker. The mixture was again heated for 30 minutes with constant stirring. The content of the beaker was filtered through the Buchner funnel and washed several times with hot water until it was free from sodium hydroxide. Finally, the residue was washed twice with 95% methanol, then quantitatively transferred into a porcelain crucible and dried at 100°C. The weight of the dry residue was noted, and the residue ignited in a furnace at 550°C. The weight of the ash left after ignition was also noted. The crude fibre content was determined from the loss in weight of crucible and its content after ignition.

2.9.5 Crude protein estimation (6.25 x N) micro kjeldahl method

One grams of the sample powder was measured for weight into 50 mL digestive Kjeldahl flask. About 20 mL of concentrated H₂SO₄, 1 tablet of Kjeldahl catalyst and a pinch of antibumbing chips were included. Same mixture sample was incinerated into a slowly boiling digestion rock, then subjected to strong heating till the digest appeared clear, before heating for further 3 hours. The digest at this point was removed and allowed to get cold, then certain amount of a known quantity was transferred into 100 mL volumetric flask up to a required mark. The Erlenmeyer flask with 100 mL of boric acid solution indicator was placed on the tip of the condenser unit of the distillation apparatus (which had been steam washed), so that the condenser tip is extended below the upper layer of the solution. Then 10 mL of the digest sample was put into the dums sample tube and made to undergo steam heating. About 10 mL NaoH solution at 40% was included in the digest and steam distilled into the Erlenmeyer flask until the content increased more than double its original quantity. As the ammonia distilled into the boric acid indicator solution, it transformed into green. A black determination was conducted in the same manner as highlighted above, exception that here, the digestion sample was substituted by 0.1 ml of distilled H₂O. The sample inside the Erlenmeyer flask was subjected to titration with 0.1 N HCl to arrive at pink end. Percentage protein was calculated as shown below:

\[
\% \text{protein} = 10 \times \left(\frac{\text{MI HCI (test)} - \text{MI HCI (Blank)}}{1000} \times \frac{10}{11} \times 6.25 \times \frac{100}{0.1}\right)
\]

2.10 Determination of Growth Performance

All growth performance parameters were calculated following the method stated by [25] as follows:
2.10.1 Weight gain (g)
This was given as final weight \( (W_2) \) – initial weight \( (W_1) \)

2.10.2 Length gain (cm)
This was given as final length \( (L_2) \) – initial length \( (L_1) \)

2.10.3 Growth Rate (GR)
This was evaluated as final weight \( (W_2) \) – initial weight \( (W_1) \) / Number of days

2.10.4 Specific growth rate (SGR)
This was evaluated as the percentage of weight gain per day
\[
SGR = \frac{\ln(W_2) - \ln(W_1)}{(T_2 - T_1)} \times 100
\]
Where:
\( W_2 \) = Weight (final) at end of time \( T_2 \)
\( W_1 \) = Weight (initial) at beginning of time \( T_1 \)
\( \ln \) = natural base of logarithm

2.10.5 Mean growth rate (MGR)
This was calculated as average weight gain in million grams per day.
\[
M.G.R = \frac{1000(W_2 - W_1)}{0.5(W_2 + W_1)t} \text{ mg/g day}
\]
Where:
\( W_2 \) = Final weight (g)
\( W_1 \) = Initial weight (g)
\( t \) = Duration of experiment in days

2.10.6 Percentage weight gain (PWG)
This was calculated as follows;
\[
PWG = 100 \frac{W_2 - W_1}{W_1}
\]
Where,
\( W_2 \) = weight (final)
\( W_1 \) = weight (initial)

2.11 Determination of Food Utilization
All food utilization parameters were calculated according to the method described by [25].

2.11.1 Food consumed (g)
This was given as 5% X body weight X No. of days

2.11.2 Food conversion ratio
This was stated as: Consumed feed (g)/ Weight gain (g)

2.11.3 Food conversion efficiency
This was given as: [Weight gain (g)/Feed consumed (g)] X 100

2.12 Statistical Analysis
Data obtained were subjected to descriptive analysis (mean and standard deviation). One way analysis of variance (ANOVA) was also used to test for the significance of difference between physico-chemical parameters of water, growth performance and food utilization parameters of each treatments group. One way ANOVA was also used to test for the significance of difference in proximate composition between the 3 treatments diets using Version 20 of predictive Analytical Software (PASW) and Microsoft Excel 2013 at 0.05 level of significance and at their relevant degree of freedom.

3. RESULTS

3.1 Proximate Composition of Experimental Diet
The summary of the mean proximate composition of the experimental diet is shown in Table 1. Mean proximate analysis of the dry matter (mg/100g) of the three experimental diets showed that crude protein content was highest \((40.61 \pm 0.13\%)\) in diet A (Coppens), followed by diet C (chicken offal based diet) (COBD) with \((38.15 \pm 0.16\%)\) and least \((37.00 \pm 0.32\%)\) in diet B (shrimp based diet) (SBD). Mean ether extract was highest in diet A \((11.71 \pm 0.10\%)\), followed by diet C \((10.00 \pm 0.30\%)\) and least in diet B \((6.70 \pm 0.12\%)\). Mean crude fibre was also highest in diet A \((7.43 \pm 0.01\%)\), followed by diet C \((5.13 \pm 0.13\%)\), and least in diet C \((4.30 \pm 0.33\%)\). Also, mean ash content was maximum in diet A \((9.10 \pm 0.12\%)\), followed by diet B \((6.67 \pm 0.33\%)\), and least in diet C \((5.00 \pm 0.00\%)\). Mean moisture content was highest in diet B \((17.37 \pm 0.31\%)\), followed by diet C \((14.84 \pm 0.14\%)\) and
least in diet A (8.50 ± 0.21%). Mean nitrogen free extract (NFE) was greater in diet C (27.76 ± 0.56%), followed by diet B (27.13 ± 0.23%) and least in diet A (22.68 ± 0.13%) (Table 1).

From the statistical point of view, nutritional composition varied significantly between the 3 feeds at p<0.05.

3.2 Growth Performance of Fish Fed Experimental Diets

The summary of the growth performance of fish fed 3 different diets are shown in Table 2. Growth performance of juveniles (*C. gariepinus*) that were administered the 3 test diets was evaluated using indices including: weight gain (g), growth rate (g/day), specific growth rate (%/day) and mean growth rate (mg/day). Weight gain (g) was highest (17189.33 ± 506.61 g) in fish fed diet A (Coppens feed), followed by 15045.33±202.42 g for fish fed diet B (shrimp based diet) and least (14357.33 ± 108.25 g) in fish fed diet C (chicken offal based diet). The Growth rate (GR) was equally highest in juveniles fed with Coppens feed (111.62 ± 3.29 g/day), followed by 97.69±1.31 g/day for fish fed diet B, and least (93.23±0.70 g/day) in fish fed diet C (Table 2).

Specific growth rate (SGR) observed was highest in juveniles fed Coppens feed (2.02 ± 0.02 %/day), followed by 1.94±0.01 %/day for fish fed diet B, and least (1.91±0.01 %/day) in fish fed diet C. Mean growth rate maintained the same line of increase in juveniles fed diet A (Coppens) reflecting highest value (11.88 ± 0.29 mg/day), followed by 11.74±0.15 mg/day for fish fed diet B, and least (11.69±0.09 mg/day) in fish fed diet C (Table 2).

Statistically, the weight gain, growth rate and specific growth rate varied significantly between the juvenile fish group fed the 3 diets at p<0.05, while the mean growth rate did not vary significantly between the juvenile fish group fed the 3 diets at p>0.05 (Table 2).

Table 1. Mean proximate composition of the experimental diets

<table>
<thead>
<tr>
<th>Indices</th>
<th>Diet A (Control) (coppens)</th>
<th>Diet B (SBD)</th>
<th>Diet C (COBD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crude Protein (%)</td>
<td>40.61 ± 0.13&lt;sup&gt;a&lt;/sup&gt;</td>
<td>37.00 ± 0.32&lt;sup&gt;b&lt;/sup&gt;</td>
<td>38.15±0.16&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Ether Extract (%)</td>
<td>11.71±0.10&lt;sup&gt;a&lt;/sup&gt;</td>
<td>6.70±0.12&lt;sup&gt;b&lt;/sup&gt;</td>
<td>10.00±0.30&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Crude Fibre (%)</td>
<td>7.43±0.01&lt;sup&gt;a&lt;/sup&gt;</td>
<td>5.13±0.13&lt;sup&gt;b&lt;/sup&gt;</td>
<td>4.30±0.33&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Ash (%)</td>
<td>9.10±0.12&lt;sup&gt;a&lt;/sup&gt;</td>
<td>6.67±0.33&lt;sup&gt;b&lt;/sup&gt;</td>
<td>5.00±0.00&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Moisture (%)</td>
<td>8.50±0.21&lt;sup&gt;a&lt;/sup&gt;</td>
<td>17.37±0.3&lt;sup&gt;b&lt;/sup&gt;</td>
<td>14.84±0.14&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>NFE (%)</td>
<td>22.68±0.13&lt;sup&gt;a&lt;/sup&gt;</td>
<td>27.13±0.23&lt;sup&gt;b&lt;/sup&gt;</td>
<td>27.76±0.56&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>*SBD = Shrimp-based diet, COBD = Chicken Offal-based diet, NFE = Nitrogen Free Extract</sup>

Values with different superscript are significantly different at p<0.05

Table 2. Mean growth performance indices of *C. gariepinus* fed experimental diets

<table>
<thead>
<tr>
<th>Growth indices</th>
<th>Diet A (Control) (coppens)</th>
<th>Diet B (SBD)</th>
<th>Diet C (COBD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial Weight (g)</td>
<td>800.00 ± 0.00</td>
<td>800.00 ± 0.00</td>
<td>800.00 ± 0.00</td>
</tr>
<tr>
<td>Final Weight (g)</td>
<td>17989.33 ± 506.61&lt;sup&gt;a&lt;/sup&gt;</td>
<td>15845.33 ± 202.42&lt;sup&gt;b&lt;/sup&gt;</td>
<td>15157.33 ± 108.25&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Weight Gain (g)</td>
<td>17189.33 ± 506.61&lt;sup&gt;a&lt;/sup&gt;</td>
<td>15045.33±202.42 g</td>
<td>14357.33 ± 108.25&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Initial Length (cm)</td>
<td>9.07 ± 0.34</td>
<td>9.11 ± 0.17</td>
<td>9.28 ± 0.27</td>
</tr>
<tr>
<td>Final Length (cm)</td>
<td>40.85 ± 0.39&lt;sup&gt;a&lt;/sup&gt;</td>
<td>39.76 ± 0.10&lt;sup&gt;b&lt;/sup&gt;</td>
<td>38.69 ± 0.35&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Length gain (cm)</td>
<td>31.78 ± 0.22&lt;sup&gt;a&lt;/sup&gt;</td>
<td>30.64 ± 0.21&lt;sup&gt;a&lt;/sup&gt;</td>
<td>29.42 ± 0.25&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Growth Rate</td>
<td>111.62 ± 3.29&lt;sup&gt;a&lt;/sup&gt;</td>
<td>97.69 ± 1.31&lt;sup&gt;b&lt;/sup&gt;</td>
<td>93.23 ± 0.70&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>SGR (%/day)</td>
<td>2.02 ± 0.02&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.94 ± 0.01&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.91 ± 0.01&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>MGR (mg/day)</td>
<td>11.88 ± 0.29&lt;sup&gt;a&lt;/sup&gt;</td>
<td>11.74 ± 0.15&lt;sup&gt;a&lt;/sup&gt;</td>
<td>11.69 ± 0.09&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>*SBD = Shrimp-based diet, COBD = Chicken Offal-based diet, SGR= Specific growth rate, MGR= mean growth rate</sup>

Values are in mean ± standard deviation

Values with different superscript are significantly different at p<0.05
3.3 Food Utilisation of Fish Fed Experimental Diets

The summary of the food utilisation indices of *C. gariepinus* juveniles fed 3 diets is shown in Table 3. Fish fed Coppens feed consumed more food (41650.00 ± 315.34g), followed by fish fed diet B with 39034.24 ± 86.34g and least in fish fed diet C with 38276.00 ± 432.97g. Juveniles fed Coppens feed did better, with a conversion ratio (2.42 ± 0.06), followed by 2.59±0.04 for fish fed diet B and 2.67 ± 0.03 for fish fed diet C. Food conversion efficiency was greater in juveniles fed diet A (41.26±0.94 %), followed by 38.54 ± 0.54 % for fish fed diet B and least (37.52 ± 0.51 %) in fish fed diet C (Table 3).

The food consumed and food conversion ratio varied significantly between the fish fed the 3 diets at p<0.05, while food conversion efficiency was not significantly different between the 3 treatment groups at p>0.05 (Table 3).

3.4 Physico-chemical Parameters of Culture Water

The summary of the mean physico-chemical parameters of culture water in tank with fish fed 3 diets is shown in table 4. For the fish group fed coppens (Diet A), the temperature of the water ranged from 27.27 – 32.83 °C, with a mean and standard deviation of 29.97 ± 0.309 °C, while pH ranged from 6.87 – 7.40, with a mean and standard deviation of 7.08 ± 0.163. The dissolved oxygen (DO) ranged from 3.37 – 5.34 mg/L, with a mean and standard deviation of 4.188 ± 0.603 mg/L, while the ammonia level ranged from 0.00 – 0.20 mg/L, with a mean and standard deviation of 0.126 ± 0.045 mg/L (Table 4).

For the fish group fed shrimp based diet (SBD) (Diet B), the temperature of the water ranged from 27.33 – 33.46 °C, with a mean and standard deviation of 30.09 ± 0.380 °C, while pH ranged from 6.91 – 7.19, with a mean and standard deviation of 7.085 ± 0.088. The dissolved oxygen (DO) ranged from 3.37 – 5.34 mg/L, with a mean and standard deviation of 4.188 ± 1.370 mg/L, while the ammonia level ranged from 0.00 – 0.20 mg/L, with a mean and standard deviation of 0.130 ± 0.052 mg/L (Table 4).

For fish fed Diet C, the temperature of the water ranged from 27.30 – 33.10 °C, with a mean and standard deviation of 30.11 ± 0.278 °C, while pH ranged from 6.93 – 7.27, with a mean and standard deviation of 7.089 ± 0.119. The dissolved oxygen (DO) ranged from 3.76 – 5.34 mg/L, with a mean and standard deviation of 4.574 ± 0.559 mg/L, while the ammonia level ranged from 0.00 – 0.17 mg/L, with a mean and standard deviation of 0.126 ± 0.045 mg/L (Table 4).

The water temperature, pH, DO and ammonia level of culture water with fish fed the 3 diets were all within the range suitable for a healthy living of fish. Statistically, the temperature, pH, DO and ammonia levels varied significantly between the culture water with fish fed the 3 diets at p<0.05 (Table 4).

4. DISCUSSION

The primary objectives of the aquaculture industry are to optimize growth and make available high-quality fish [2]. The increase in the World-wide awareness of fish as a treasured source of protein, has led to an increase in the prospects of aqua feeds with diets being specially design to meet the nutritional needs of species, life cycle and the health conditions of fish [3]. Fishmeal has been the most used source of protein incorporated in culture fish diets, despite the fact that, fish supply is not increasing globally [4]. In fish farming, feed and its application still remain a limiting constraint to fish

<table>
<thead>
<tr>
<th>Growth Indices</th>
<th>Diet A (Control) (coppens)</th>
<th>Diet B (SBD)</th>
<th>Diet C (COBD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Weight Gain (g)</td>
<td>17189.33 ± 506.61ᵃ</td>
<td>15045.33 ± 202.42ᵇ</td>
<td>14357.33 ± 108.25⁷</td>
</tr>
<tr>
<td>Food Consumed (g)</td>
<td>41650.00 ± 315.34ᵃ</td>
<td>39034.24 ± 86.34ᵇ</td>
<td>38276.00 ± 432.97⁷</td>
</tr>
<tr>
<td>FCR</td>
<td>2.42 ± 0.06ᵃ</td>
<td>2.59 ± 0.04ᵇ</td>
<td>2.67 ± 0.03⁷</td>
</tr>
<tr>
<td>FCE (%)</td>
<td>41.26 ± 0.94ᵃ</td>
<td>38.54 ± 0.54ᵇ</td>
<td>37.52 ± 0.51ᵃ</td>
</tr>
</tbody>
</table>

*ᵃSBD = Shrimp-based diet, COBD = Chicken Offal-based diet
ᵇFCR= Food conversion ratio,
ᶜFCE = Food conversion efficiency
Values are in mean ± standard deviation
Values with different superscript are significantly different at p<0.05
had the slowest growth. This can be attributed to while fish fed chicken offal based diet (COBD) faster than fish fed shrimp based diet (SBD), by fish as reflected in the proximate levels of the experimental diets in terms of nutrient required an indicator of the completeness of the three observed in this study for a period of 22 weeks is met by that feed. Growth response for fish how efficient an animal's nutrient requirement is driven towards developing feeds using locally available ingredients and some common waste products such as chicken offals and shrimp waste within the Calabar environments as protein sources, while trying to compare it with Coppens feed which over time has been proven to be of good quality, giving a better yield in terms of growth.

The nutritional composition of the 3 feeds used differed in this study, with coppens being the best from the nutritional point of view, but all 3 feeds were well formulated to have a balanced diet. The differences in the nutritional levels of the 3 diets could be due to the difference in the components of the 3 diets. This corroborated with the findings of [20] and [21], who both purported that, the nutritional constituents of fish feed which over time has been proven to be of good quality, giving a better yield in terms of growth.

The importance of the amount of feed consumed cannot be over emphasized, because of its vitality for checking the ratio of food conversion (FCR) and food conversion efficiency (FCE). Since feed are costly, FCR or FCE are important factors for the accurate use of feed [28]. Research has consistently highlighted that good knowledge of FCR helps the farmer to satisfactorily feed the fish [29,30,31,32,33,34, 27]. In this study, Diet A were more readily converted by fish as indicated by the high FCR and FCE, followed by Diet B and then Diet C as the least. The high conversion of coppens could be due to it having a strong fish odour, as well as being more nutritionally balanced than the other diets.

The water quality parameters varied between treatment tanks, although all water quality parameters monitored were normal and at a tolerable limits for the rearing of {C. gariepinus}.
5. CONCLUSION

In connection with these findings, it was concluded that shrimp based diet (SBD) and chicken offal based diet (COBD) competed positively with coppens regarding nutritional composition, growth performance and feed utilisation of C. gariepinus. Despite the fact that coppens yielded better nutritional composition, fish growth performance and feed utilisation, fish fed SBD and COBD to maturity can be used as a reliable broodstock. In conclusion, the use of coppens feed for catfish farming is advocated, but for a more cost effective system, a formulated diet using shrimp or chicken offal as the animal protein source is recommended to fish farmers in Nigeria. More researches should be carried out on using chicken offals and shrimp based diets in fish feed formulation.

ETHICAL CONSIDERATION

The authors ensured that all ethical and other basic principles underlying behavior and advancing welfare for the use of animals in research, including handling, relevant laws and regulations were considered before proceeding with the research. Permission was also received from the relevant bodies for the use of fish for this experiment.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

18. Glencross BD, Booth M, Allan GL. A feed is only as good as aquaculture feeds. Aquaculture Nutrition. 2007;13:17-34.

© 2018 Ayim et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
http://www.sdiarticle3.com/review-history/43149