Isolation and Antifungal Effects of Plants Extracts on *Malassezia* species Isolated from Scalps of Primary School Pupils and Bingham University Students

Maikenti James Ishaku¹, Egah Ruth Grace², Adogo Lillian Yami² and Koggie Amos Zamfara²

¹Department of Zoology, Federal University of Lafia, Nasarawa State, Nigeria.  
²Department of Biological Sciences, Bingham University, Nigeria.

Authors' contributions

This work was carried out in collaboration among all authors. Author MJI designed the methodology and supervised the study. Authors KAZ and ERG managed the analysis of the study and the literature of the study. Author ALY prepared the first draft of the manuscripts. All authors read and approved the final manuscript.

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ABSTRACT

**Aim:** The aim of the research was to evaluate the antifungal activities of the leaf extracts of *Senna alata* and *Lawsonia inermis* on *Malassezia* spp. isolated from the scalps of primary school pupils and Bingham University students.  

**Place and Duration of the Study:** This study was carried out in the department of Biological sciences, Bingham University Karu Nasarawa state between July to September, 2017.  

**Methodology:** The plant species *Senna alata* and *Lawsonia inermis* was collected from Garki district of the Federal Capital Territory Abuja and Sayina village of Auta-balefi Karu Local Government Area of Nasarawa state respectively. These plant species were identified by plant experts in the Department of Biological Sciences, Bingham University, using characteristic features of the leaves such as the shapes, sizes and flower corresponding to the herbarium specimens.

*Corresponding author: E-mail: jamesmaikenti@yahoo.com;*
### INTRODUCTION

Dandruff is a common scalp condition affecting majority of the population at pre-pubertal and post pubertal age, cutting across ethnicity, race and gender [1]. It is also believed to be caused by both microbial and non-microbial factors, with the most accepted etiological agent to be a lipophilic yeast that is of the genus *Malassezia* [2]. These etiologic agents are prevalent in regions with sebaceous glands because they are lipid dependent [3].

The Lipophilic dimorphic yeast *Malassezia furfur* is one of the common causative agents of dandruff including *Malassezia restricta* and *Malassezia globosa*. *Malassezia furfur* feeds on the dermal lipids and proteins which enhances and facilitates its lipase activity, causing inflammation and dermal tissue damage [2].

Dandruff cannot be completely eliminated, but can only be managed or controlled [4,5]. Eradication of dandruff causing fungus will facilitate effective treatment of dandruff leading to its management and effective control. Many substances available for the treatment of dandruff consist of various chemical based antifungal agents like Clotrimazole, Amphotericin B, Zinc pyrithione, Salicylic acid, Imidazole derivatives, Glycolic acid, Steroids, Sulfur, Tar derivatives and Nystatin etc. and are mostly associated with side effects such as dryness of hair/skin, associated cytostasis, and eczema, and also frequent reoccurrence made therapy costlier. So, it is imperative to search for drugs that are safe, cost-effective, and eco-friendly [6].

Plants rich in a wide array of antifungal secondary metabolites are gaining importance as a primary source of commercial medicines and drug leads, and have been used for the treatment of skin disorders for centuries [7,8]. Herbal drug technology has picked up strongly in terms of the extraction and characterization of active compounds and also the processing of herbs into medicine, considering the fact that modern medicine is not capable of providing a
"cure all" solution for the pathogenic infections. Herbal therapy is considered as a therapeutic alternative, as a safer choice than synthetic medicines, or sometimes even as the only successful therapeutic way left to treat these disorders [6]. Therefore, this study was designed to isolate and determine the antifungal activities of *Senna alata* and *Lawsonia inermis* leaf extracts on *Malassezia* spp.

Phytochemical analyses revealed that *Senna alata* leaves contained saponins, alkaloids, flavonoids, tannins, phenols anthraquinone, protein and carbohydrate (glycosides) [9]. While *Lawsonia inermis* contain naphthalene derivatives, quinoids, β-sitosterol glycoside, xanthones, flavonoids, gallic acid, coumarins and lawsoniasides. Lawsone, 2-hydroxy-1, 4-naphthoquinone is responsible for henna’s fungicidal activity.

2. MATERIALS AND METHODS

2.1 Study Area

This study was conducted at Auta-Balefi. Two sites were considered, Bingham University and Kodope primary school. Bingham University and Kodope Primary School are both located in Auta-Balefi, along KM 26 Abuja Keffi Expressway Kodope, Karu, Nasarawa state. Bingham University alongside Kodope primary school covers a land mass of 200-220 square meters, it has a population of about 2000 students and is geographically located at latitude 8º 50º N and longitude 7º 52º E. The University was established in the year 2005 [10]. Kodope Primary school is located at Bingham University Junction.

2.2 Collection and Identification of Plant Samples

Fresh leaves of the two plants used in this study were collected from Area one, Garki Abuja and Sayina village Auta-Balefi Karu LGA Nasarawa State. These leaves were collected separately into sterile polythene bags and transported immediately to the laboratory for identification and processing. The plants were identified by expert in the department of Biological Sciences Bingham University Karu using herbarium key (ER135) for *Senna alata* (*Cassia alata*), and key (27544) for *Lawsonia inermis*. Characteristic features of the leaves such as shapes, sizes and flower such as the colour were used in the identification of plants. Other features include plant height, fruits they produce, nature of the branches, colour of the stems, present or absence of spines, and number of flowers.

2.3 Preparation of Extracts

The samples previously collected were dried at room temperature separately in the laboratory for one week. The dried leaves of *Senna alata* and *Lawsonia inermis* were further dried in a hot air oven separately to make grinding easier; they were grounded mildly into coarse powder using clean mortar and pestle after which it was transferred into a sterile bijou bottles and stored at -20º in a refrigerator before use [11].

2.3.1 Ethanol extracts

Twenty grams of the powdered material was extracted with 100ml of 80% ethanol, with constant agitation for 3 days and then filtered using a muslin cloth (Vandee and Peeranuch 2009).

2.3.2 Aqueous extraction

2.3.2.1 Boiling

Ten grams of each of the plant powdered samples were boiled in 100 ml of water in separate beakers for 30 mins. The resulting extracts were filtered using Whatman No. 2 filter paper [12].

2.3.2.2 Soaking

Ten grams of the plant powdered sample was soaked in 100ml of distilled water for 24hours. The resulting extracts was allowed to cool down and then filtered using whatman No. 2 filter paper [12].

2.4 Collection and Isolation of the Test Organisms

Dandruff samples were obtained by scrapping the scalp surface with sterile combs. The combs used were sterilized by washing in detergent and soaking in 10% bleach solution to prevent contamination. Some portion of the scrapping from the scalp was inoculated over the surface of modified Dixon agar slants, transferred to the laboratory on the same day and inoculated at 30ºC for 14 days. The morphology was examined on leeming and Notman agar after incubation at 32ºC for 7 days [13].
2.5 Identification of Fungus

Identification of *Malassezia* was made microscopically by Löffler’s Methylene Blue Staining and a drop of 10% potassium hydroxides (KOH). Each slide was examined under high power field of microscope (40X objective) to reveal the presence of hyphae and budding yeast cells which generally exhibit the characteristic appearance of “spaghetti and meatballs” [14,15].

2.6 Biochemical Tests

The gross morphology of suspected colonies of *Malassezia* which shows globose blastoconidia and mycelium was noted prior to biochemical test. Different biochemical tests such as the catalase test using hydrogen peroxide, bile Esculin splitting test, and Tween 20 and 80 utilization tests were carried out.

2.6.1 Catalase test

A small amount of fungal colony was transferred to a surface of a clean, dry glass slide using a wire loop, followed by a drop of hydrogen peroxide. Production of gas bubbles on adding a drop of hydrogen peroxide indicated a positive reaction.

2.6.2 Bile esculin splitting test

A fungal colony was streaked on bile Escculin agar and incubated at 37°C for 2-5 days and development of black colored byproducts of Escculin hydrolysis indicates a positive test [16].

2.6.3 Tween utilization tests

A single colony from the 21 cultured sample plates was picked using a sterile cotton swap to make a lawn culture on the SDA plates and three wells were made using a borer on to which Tween 20 and 80 were inoculated. Utilization of Tween compounds was assessed by the degree of growth and or reaction (precipitation) of the lipophilic yeast around individual well [16].

2.7 Evaluation of Antifungal Activity of Plant Extracts

The antifungal effect of the plant extracts was evaluated by agar well diffusion method [14]. The surface of Sabouraud Dextrose Agar plates was inoculated with test microorganisms by spreading the surface of the media using sterile swabs. Three wells were made using the borer with diameter 8.00 mm each for *Senna alata* and *Lawsonia inermis* extracts as well as the control (ethanol) to accommodate the three technical replicates. 10µl of the extracts was dispensed in the wells. The plates were incubated at 37°C for 2-5 days. The antifungal activity was determined by calculating the mean zones of inhibition of the 21 biological samples.

3. RESULTS

The overall prevalence of *Malassezia* spp. infection in the study area is 25.3% as shown in Table 1. The study also isolated three species as follows, *Malassezia restricta* 11(52.3%), *Malassezia furfur* 5(23.8%) and *Malassezia globosa* 5(23.8%), out of which *Malassezia restricta* was most prevalent. *M. restricta* was the main species causing dandruff, as it was isolated from 52% of the dandruff samples cultured; followed by *M. furfur* (24%) and *M. globosa* (24%) while *M. pachydermatis* and *M. obtusa*. *Malassezia slooffiae* and *M. sympodialis* were not found in any of the cases.

Table 2 shows the relationship of *Malassezia* spp. to gender of the students to be 7(33.3%) for males and 14(66.6%) for females (P = 0.367).

Table 3 shows higher occurrence among age group 21-25 (61.7%) followed by age group 16-20 (33.2%). *M. restricta* and *M. globosa* were not isolated among age group 10-15. *M. furfur* was found in all age group. The results did not demonstrate a significant relationship between the frequency of *Malassezia* species and age (p = 0.187), as well as gender (P = 0.367).

Table 4 shows the prevalence of *Malassezia* species with respect to institutions where samples were collected (p=0.043). Out of the samples collected from Kodope primary school only one was positive for *Malassezia furfur*. *M. restricta* and *M. globosa* were not isolated from the primary school pupils. Samples collected from Bingham university students expressed all the three species isolated in the study.

Table 5 shows the mean zones of inhibition in millimetres and mean standard error (SEM) observed upon the application of aqueous extracts (soaked and boiled) and ethanolic extracts of *L. inermis* and *S. alata*, with ethanol serving as a control. The mean zone of inhibition for soaked extracts of *L. inermis* did not show any inhibitory effect but *S. alata* had inhibitory
effects on the isolates. Also, both plants species extracted through boiling did not have any inhibitory effect. Ethanolic extract of S. alata had higher zone of inhibition of 11.00±0.969 compared to the ethanolic extract of L. inermis (7.71±0.876).

Table 1. The prevalence of Malassezia species in the study area

<table>
<thead>
<tr>
<th>Study locations</th>
<th>No. examined</th>
<th>No. positive</th>
<th>No. negative</th>
<th>Prevalence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kodape</td>
<td>23</td>
<td>1</td>
<td>22</td>
<td>25.3%</td>
</tr>
<tr>
<td>Bingham</td>
<td>60</td>
<td>20</td>
<td>40</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>83</td>
<td>21</td>
<td>62</td>
<td></td>
</tr>
</tbody>
</table>

Table 2. Malassezia species isolated in relation to gender

<table>
<thead>
<tr>
<th>Species</th>
<th>Males</th>
<th>Females</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Malassezia restricta</td>
<td>5(23.8)</td>
<td>6(28.5)</td>
<td>11(52.3)</td>
</tr>
<tr>
<td>Malassezia furfur</td>
<td>1(47)</td>
<td>4(19)</td>
<td>5(23.8)</td>
</tr>
<tr>
<td>Malassezia globosa</td>
<td>1(4.7)</td>
<td>4(19)</td>
<td>5(23.8)</td>
</tr>
</tbody>
</table>

Table 3. Malassezia species in relation to age

<table>
<thead>
<tr>
<th>Organism</th>
<th>10-15</th>
<th>16-20</th>
<th>21-25</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Malassezia restricta</td>
<td>0(0)</td>
<td>4(19)</td>
<td>7(33.5)</td>
<td>11(52.3)</td>
</tr>
<tr>
<td>Malassezia furfur</td>
<td>1(4.7)</td>
<td>1(4.7)</td>
<td>3(14.2)</td>
<td>5(23.8)</td>
</tr>
<tr>
<td>Malassezia globosa</td>
<td>0(0)</td>
<td>2(9.5)</td>
<td>3(14.2)</td>
<td>5(23.8)</td>
</tr>
<tr>
<td>Total</td>
<td>1(4.7)</td>
<td>7(33.2)</td>
<td>21(100)</td>
<td>13(61.7)</td>
</tr>
</tbody>
</table>

Table 4. Malassezia species isolated in relation to institution

<table>
<thead>
<tr>
<th>Organism</th>
<th>Kodape pri. sch.</th>
<th>Bingham university</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>M. restricta</td>
<td>0(0)</td>
<td>11(52.3)</td>
<td>11(52.3)</td>
</tr>
<tr>
<td>M. furfur</td>
<td>1(4.7)</td>
<td>4(19)</td>
<td>5(23.8)</td>
</tr>
<tr>
<td>M. globosa</td>
<td>0(0)</td>
<td>5(23.8)</td>
<td>5(23.8)</td>
</tr>
<tr>
<td>Total</td>
<td>1</td>
<td>20</td>
<td>21</td>
</tr>
</tbody>
</table>

Table 5. Mean standard error (SEM) of different extraction methods of (Senna alata and Lawsonia inermis) on Malassezia sp.

<table>
<thead>
<tr>
<th>Methods of extraction</th>
<th>Plants sp.</th>
<th>Soaking</th>
<th>Ethanolic extracts</th>
<th>Control(Ethanol)</th>
<th>Boiling</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Mean(SEM)</td>
<td>Mean(SEM)</td>
<td>Mean(SEM)</td>
<td>Mean(SEM)</td>
</tr>
<tr>
<td>S. alata</td>
<td>10.00±0.969</td>
<td>11.00±0.969</td>
<td>6.00±0.756</td>
<td>0.00±0.00</td>
<td></td>
</tr>
<tr>
<td>L. inermis</td>
<td>0.00±0.00</td>
<td>07.71±0.876</td>
<td>4.000±0.396</td>
<td>0.00±0.00</td>
<td></td>
</tr>
</tbody>
</table>
Plate 3. Culture of Malassezia sp.

4. DISCUSSION

Malassezia species are yeasts that make up part of the microbiota of the mammalian skin. They are prevalent in regions with sebaceous glands due to their dependency on lipids [3]. This study revealed 25.3% prevalence of Malassezia species. The low level of infection recorded in this study could be due to use of other synthetic antifungal creams by the subjects. This prevalence is low compared to the findings of Hedayati et al. [17] and Zareei et al. [18] both of whom worked independently and reported higher prevalence of 70% and 93.5% respectively in their studies.

Three species that characterised this study were M. restricta, M. furfur and M. globosa. Out of the three species encountered in this study, M. restricta was the most common species followed by M. furfur and M. globosa. Other similar studies who reported these species were the reports of Shivaprakas et al. [19], Shu’aibu et al. [14], and Zareei et al. [18]. However, their findings differ slightly with this finding in that they all reported different species to be the most prevalent. This is evident in the reports of Zareei et al. [18] who reported M. globosa as the most significant species while M. sympodialis (6.5%) and M. slooffiae (0.8%) were the least fungal species isolated. Hedayati et al. [17] reported the M. globosa as the most commonly isolated species followed by M. furfur and M. restricta. Shu’aibu et al. [14] also reported M. globosa as the most predominant species. These variations may be due to the fact that Malassezia species implicated with dandruff vary at different geographical locations, study participants or isolation method used.

Higher infection was recorded among University students with all the three species isolated from the students. This finding disagrees with the finding by Shivaprakash et al. [19] who recorded higher prevalence among age group 10-19. This is so because other studies indicated differences in the colonisation pattern of Malassezia species with different age groups with higher colony counts in adults than in children and a decrease in the elderly individual [20]. This condition may be due to high sebum production in the above-mentioned age groups, microbial metabolism or susceptibility of the individual.

This study evaluated the anti-malassezia effects of the two plants species S. alata and L. inermis. The result reveals S. alata to exhibit some antimalassezia effect on the isolates with mean zone of inhibitions of 11± 0.969 for ethanolic extracts and 10± 0.969 for soaked (aqueous) extracts. From these findings and other reports from Borade et al. [7], Jung et al. [21] and Wandee and Peeranuch [11], the ethanolic extracts seem to exhibit higher zone of inhibition than the aqueous solution. This may not be far from the reports of Natarajan et al. (2005) that different solvents have different solubility capacities for different phyto-constituents, thus, the differences in the activities of the various extracts [20,21,22].

In the case of L. inermis plant extract, the result shows lower anti-malassezia effect compared to S. alata with no zone of inhibition for soaked and boiled extracts. Only the Ethanolic extracts of
5. CONCLUSION

This study reveals that naturally acquired herbs of *S. alata* especially and *L. inermis* are effective in combating *Malassezia* species which are the causative agents of dandruff and hence, the use of naturally acquired herbs should be encouraged in combating dandruff.

CONSENT AND ETHICAL APPROVAL

Ethical approval to conduct research was sought from Nasarawa State Hospital Management Board (NSHMB) through the Research Ethics Committee of Nasarawa State Ministry of Health Lafia, in line with the guidelines required for conducting research on human samples. Further Approval was sought from the respective school management as well as Informed consent from the volunteers and their parents / guardians.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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