Modelling of Biphasic Biogas Production Process from Mixtures of Livestock Manure Using Bi-logistic Function and Modified Gompertz Equation

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Authors’ contributions

This work was carried out in collaboration among all authors. Author CCO designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors CON and CEN managed the analyses of the study. Authors CCO and NAN managed the literature searches. All authors read and approved the final manuscript.

ABSTRACT

In this study, anaerobic digestion (AD) of three livestock manure: (poultry manure (PM), pig dung (PD), and cow dung (CD) was conducted at different mixed ratios under mesophilic (25-35°C) conditions. Two kinetic models, the modified Gompertz and bi-logistic function model were used to simulate the cumulative biogas yield from the experiments, and model parameters simultaneously obtained. The biogas production profile appeared diauxic-like or biphasic with multiple peaks, revealing the complexity and multi-component nature of the substrates. There was an increase in biogas yield from the treatments, PD/CD 1:1; 37.25 dm³ (3.47%), PD/CD 3:1; 38.41 dm³ (6.96%), CD/PM 1:1; 26.76 dm³ (16.80%) and CD/PM 3:1 24.31 dm³ (6.11%), whereas PD/CD 2:1 (15.41 dm³) and CD/PM 2:1 (22.57 dm³) exhibited inhibitory effect. However, statistical analysis (ANOVA) indicated a significant difference in biogas yield from PD alone (36 dm³) compared to CD alone (22.91 dm³). The two models showed good performance in the simulation of the AD process, with high correlation coefficients, an indication of a very strong relationship between experimental data and model parameters. However, the bi-logistic function model showed a better fit in the simulation of the experimental values, as it was able to capture the curves in the plots, with a higher
correlation coefficient R² (0.9920 - 0.9985) than the modified Gompertz model (0.9797 - 0.9968). This work has shown that the phenomenon of diauxic growth in the anaerobic digestion of complex organic substrates could be captured quantitatively in the kinetic model using bi-logistic function model.

Keywords: Anaerobic co-digestion; biogas yield; diauxic growth; kinetic model; livestock manure.

1. INTRODUCTION

Given the intensified agricultural activities to address global food crises and ensure food security, there has been an increased waste generation. Production of "green energy" from agro-wastes is a panacea for the challenges (energy depletion and waste management) confronting humanity as a result of climate change and natural resource vulnerabilities. Animal husbandry produces an enormous quantity of manure that poses a serious problem for the environment. Chicken droppings are mainly made up of nitrates. Nitrate pollution is noxious because of its potential role in eutrophication, methemoglobinemia, and nitrosamines formation [1] and effective treatment methods are therefore very necessary. Anaerobic digestion (AD) has proven to be an attractive and efficient pathway to this, offering multiple advantages, such as reduced pollution and emission of greenhouse gases, eliminate pathogens, biomass reduction, stabilization of wastes, and production of biogas), which is considered a competitive renewable energy source.

The application of untreated manures has been a common practice in feeding fish in the Far East for many years [2]. The growth of planktons in fish ponds has been boosted by the introduction of poultry and other livestock manures directly into the pond, but the repulsive odour tends to have an adverse effect on the palatability of the fish. The use of commercially prepared feed in fish farming is capital intensive and it is not suitable and viable in many countries. Recycling organic wastes in fish production after composting or by fermenting the wastes in bioreactors have been an alternative method [2].

Animal manure is majorly composed of lignocellulosic fibers that were not completely digested by the animals [3]. Livestock manures are nutrient-rich soil conditioner and also a promising resource for "green energy" production by anaerobic digestion, which significantly reduces the volume and stabilizes the manure. One of the primary advantages of utilizing manures as a source of biogas production is because of their readily availability as a domestic resource in rural communities and can reduce the dependency on fossil fuels. Waste to Energy (WTE) technologies such as biogas technology, therefore should be extensively employed for the utilization of animal manure and to mitigate the climate change occasioned by the unscientific management of animal manure [4]. Anaerobic digestion (AD) of livestock manure for biogas production does not reduce its value as a soil conditioner or a fertilizer supplement because the available nitrogen, ammonium, and other mineral nutrients that remain in the digestate are considerable enough to support plant growth [5,6].

The significance of AD as an ecofriendly approach to waste management and biogas production has triggered off several research efforts on the different ways to improve process stability, biomass conversion efficiency, and enhance biogas production. Improving the compositional characteristics of substrates by anaerobic co-digestion of one substrate with another at different ratios is one of the methods that have been extensively investigated and widely adopted in enhancing biogas yield. For example, anaerobic co-digestion of animal manure with kitchen wastes [7-9], animal manure with straw [10-12], with water Hyacinth [13-16], with dedicated energy crops [17,18], animal manure with another [19-21] and sewage sludge [22] have been reported.

The microbiology/biochemical processes and operational characteristics of bio-digesters have led to the development of different designs and types of bio-digester, batch mode, sequencing batch, and continuous bioreactors. To develop an efficient and very reliable design of biodigester and assess its efficiency and performance, appropriate mathematical models describing the process are important [23]. There are several documented mathematical models such as models for calculating biogas production based on stoichiometry, and models based on reaction kinetics which also take into consideration, product inhibition, substrate limiting, etc [24,25].
Biogas production process contains several complex interconnections. The different parameters required to characterize the process complicates the development of a well intelligible model. However, several kinetics models have been developed and applied in describing anaerobic digestion process [26-30]. Several compositional analyses conducted on agro-wastes [31,32] have shown that they are highly complex and multi-component in nature, some with two or more carbon and energy sources such as lignocellulose, starch, fat, etc. During anaerobic digestion and biogas production, if two or more of these complex compounds are present in the medium, microbial growth will occur preferentially on the fast metabolizable substance. This is followed by a temporary growth cessation (another lag period) before the utilization of the others. The trade-off between fast metabolized substrate before the switch-off to the slowly metabolized substrate is known as biphasic or diauxic growth. It is a phenomenon whereby a population of microorganisms, when presented with two or more carbon and energy sources, exhibits biphasic exponential growth intermitted by a lag-phase of minimal growth [33]. The consequence of this is that biogas production will occur in phases, with periods of low and accelerated biogas yield (multiple peaks). Kinetic assessment of biphasic or diauxic-like anaerobic digestion (AD) process of livestock manures using bi-logistic function model is very rare in the literature. To fill the existing gaps in this field of study, this work investigated the feasibility of simulation of diauxic-like or biphasic biogas production process from anaerobic digestion of mixtures of livestock manures using bi-logistic function and modified Gompertz equation.

2. MATERIALS AND METHODS

Three livestock manure, poultry manure, pig, and cow dung were used in this study. The cow dung (CD) was collected from an abattoir while the pig dung (PD) and poultry manure (PM) were obtained from within the Federal University of Technology, Owerri, Imo State, Nigeria. Prior to anaerobic digestion, the samples were sun-dried, objectionable solids removed and the particle size reduced by grinding and sieving. The proximate compositions of the samples were determined by adopting standard methods [34].

2.1 Experimental Set-Up and Biogas Production

The experiment was designed such that PD/CD was co-digested at varying ratios: 1:1, 2:1, and 3:1. CD/PM were digested in like manner while CD and PD alone served as the control. Slurries of the different mixed ratios were prepared with water and subsequently fed into labeled 10L capacity bioreactors. The picture of the experimental set-up is shown in Fig. 1 and the details of the bioreactor content in Table 1. The reactors were inoculated with strained liquor of fresh cow rumen waste (inoculum), and the effective volume of 8L was achieved by adding water. After thorough mixing, the initial pH of the slurry was adjusted to 7.50 using NaOH.
Table 1. Details of bioreactor content

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Treatment Ratios</th>
<th>TS (%)</th>
<th>VS (%)</th>
<th>Inoculum (L)</th>
<th>Final Volume (L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PD/CD</td>
<td>PD/CD 1:1</td>
<td>5.88</td>
<td>4.18</td>
<td>1.6</td>
<td>8</td>
</tr>
<tr>
<td></td>
<td>PD/CD 2:1</td>
<td>5.76</td>
<td>4.28</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>PD/CD 3:1</td>
<td>5.74</td>
<td>4.33</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>PD Alone</td>
<td>5.70</td>
<td>4.48</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CD/PM</td>
<td>CD/PM 1:1</td>
<td>5.79</td>
<td>4.14</td>
<td>1.6</td>
<td>8</td>
</tr>
<tr>
<td></td>
<td>CD/PM 2:1</td>
<td>5.82</td>
<td>4.05</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>CD/PM 3:1</td>
<td>5.83</td>
<td>4.01</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>CD Alone</td>
<td>5.88</td>
<td>3.89</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

The reactors were properly sealed and the biogas outlet connected to the biogas harvesting system previously filled with water and allowed to overflow. Anaerobic digestion (AD) was under a temperature range of 25 - 35°C and the pH of the digesting slurry was read with a digital pH meter at alternate days and maintained at 7.0 - 7.80 throughout the fermentation period which lasted for 84 days. Daily biogas produced in each of the reactors, collected by the downward water displacement method was measured after manual agitation. The effect of the test parameters on anaerobic digestion was evaluated by the maximum cumulative biogas yield.

**Kinetic Modeling:** Two kinetic models, the modified Gompertz model (Equation 1), and logistic function models (Equation 2) were used to simulate the mean cumulative biogas yield obtained from the experiments. Selection of the most suitable kinetic model should not only be to predict the efficiency of particular bioreactors, but also to analyze correctly, the metabolic pathways and mechanisms involved during the anaerobic digestion [35].

\[
Y_t = Y_m \cdot \exp \left\{- \exp \left[ \frac{U \cdot e}{Y_m} (\lambda - t) + 1 \right]\right\}
\]  

Where:

\(Y_t\) = the cumulative biogas production (dm³)

\(Y_m\) = the biogas production potential (dm³)

\(U\) = the maximum biogas production rate (dm³/day)

\(\lambda\) = Lag phase period (days)

\(t\) = cumulative time for biogas production (days)

\(e\) = mathematical constant (2.718282)

The modified Gompertz equation [36] was fitted into the experimental data to predict rate of biogas with assumption that biogas production rate in batch system is a function of the specific growth rate of methanogenic bacteria in the digester. The maximum specific biogas production rate \(U\), lag phase time \(\lambda\), and biogas production potential \(Y_m\) were estimated by performing non-linear regression analysis with aid of Sigma Plot version 10.0.

Because the anaerobic digestion process mirrored diauxic growth pattern with multiple peaks of biogas production, the logistic function model [35] was modified as shown in equation 2. The equation was fitted into the experimental values, and kinetic constants generated by non-linear regression analysis the same software.

\[
y = \frac{P_{b1}}{1 + \exp \left( \frac{4R_{m1}(\lambda_1 - t)}{P_{b1}} + 2 \right)} + \frac{P_{b2}}{1 + \exp \left( \frac{4R_{m2}(\lambda_2 - t)}{P_{b2}} + 2 \right)}
\]

Where:

\(y\) is the biogas yield (dm³) with respect to time \(t\) (days)

\(P_{b1}\) is the maximum biogas potential of the substrate (dm³) before the second lag

\(P_{b2}\) is the maximum biogas potential of the substrate (dm³) in the second phase

\(R_{m1}\) is the maximum biogas production rate (dm³) before the second lag

\(R_{m2}\) is the maximum biogas production rate (dm³) in the second phase

\(\lambda_1\) is the first lag phase time (days)

\(\lambda_2\) is the second lag phase time (days)

\(t\) is the time (days).

Statistical analysis (ANOVA) of cumulative biogas yield from the different treatments was carried out using Post-Hoc Duncan test implemented in IBM SPSS statistics software version 20.0.

3. RESULTS

3.1 Proximate Composition of the Substrates

On dry weight basis, the proximate compositions of the different livestock manure are presented in
Table 2. Two very important parameters, the C/N ratio and volatile solids (VS) contents of PD are 10.00 and 68.84%; the CD contained 37.00 and 59.77%, whereas PM had 15.00 and 67.62%, respectively.

3.2 Biogas Production

The anaerobic digestion (AD) and daily biogas production profile for the different ratios of PD/CD are shown in Fig. 2. The lag phase lasted for 8 days in PD/CD 1:1 and 13 days in PD/CD 3:1, the first 14 days in PD/CD 2:1 and PD alone recorded very low non-flammable gas production. The peaks of gas production were on day 21 in PD/CD 1:1 (2390 ml); day 23 (1430 ml) and day 31 (1500 ml) in PD/CD 2:1; and day 22 (2920 ml) in PD 3:1, whereas multiple peaks were observed in the anaerobic digestion of PD alone. The flammability test indicated that the biogas became flammable on the 16th and 18th day in PD/CD:1:1 and PD/CD 3:1, whereas in PD/CD 3:1 and PD alone the biogas became flammable on the 21st day.

A similar pattern of AD and gas production was observed in CD/PM (Fig. 3). The lag period was for 11 days in CD/PM 1:1 and 5 days in CD/PM 2:1, CD/PM 3:1 and CD alone. The peak of biogas production was recorded on day 22 (2710 ml) in CD/PM; 18th (1800 ml) and 19th (1720 ml) day in CD/PM 2:1. Two peaks (19th and 22nd day) of gas production were also observed in CD/PM 3:1, with 1850 ml and 1903 ml, respectively.

The maximum cumulative biogas yield in the different ratios and the corresponding percentage increase and inhibition in biogas production are summarized in Fig. 4. PD/CD 1:1 and PD/CD 3:1 showed 3.74 and 6.96% increase in biogas yield; CD/PM 1:1 and CD/PM 3:1 was 16.80 and 6.11%, respectively. Inhibitory effects were recorded in PD/CD 2:1 and CD/PM 2:1.

Statistical analysis (ANOVA) revealed no significant difference in biogas yield in any of the treatments relative to the control, but indicated a significant difference (P ≤ 0.05) in cumulative biogas yield from PD alone compared to CD alone, and significant inhibition in PD/CD 2:1.

3.3 Kinetic Study

Plots of simulation of experimental data from the different ratios of PD/CD, CD/PM, and control with modified Gompertz model are shown in Figs. 5 and 6. The kinetic constants estimated using the non-linear regression is presented in Table 3. The anaerobic digestion process was well described by the modified Gompertz model, with a correlation coefficient (R²) > 0.98. Similarly, the experimental data were also simulated using bi-logistic function equation. The bi-logistic model fitted properly and much better than the modified Gompertz model as it captured all the curves (Figs. 7 and 8), with R² > 0.99. Due to the biphasic nature of the anaerobic digestion process, two sets of model parameters (Pb₁; Rm₁; λ₁ and Pb₂; Rm₂; λ₂) were generated, as presented in Table 4, and the correlation coefficients (R²) which indicate the goodness of fit of the models for the experiments are shown in Table 5.

4. DISCUSSION

The results of this study showed an increase in cumulative biogas yield in some of the mixtures of PD/CD and CD/PM compared to PD and CD alone. The treatments did not significantly improve biogas production relative to the control (Fig. 4); however, a significant difference in biogas production was recorded in PD alone, with 57.14% higher than CD alone. This may be among other factors, attributed to the higher volatile solids (VS) content of PD. This finding is in agreement with Olufemi et al. [37]. The results of their study revealed that co-digestion of chicken droppings and cow dung increased biogas yield as compared to pure samples of either chicken droppings or cow dung. Kaffe and Kim [38] evaluated the performance of anaerobic digesters using a mixture of apple waste (AW) and swine manure (SM). The studies were carried out using both batch and continuous digester. The results showed that mixture of AW and SM improved the biogas yield by approximately 16% and 48% at mesophilic and thermophilic temperatures, respectively, relative to SM alone, but no statistical difference was found in the methane yield.

Ambient temperature kinetic assessment of biogas production from co-digestion of horse and cow dung was evaluated by Yusuf et al. [39]. The result showed that biogas yield was optimized when horse and cow dung was mixed at a ratio of 3:1.

There was a significant inhibitory effect on biogas production in the bioreactor with PD/CD 2:1. It is apparent from these results and previous studies [40], that the improvement of biogas yield by co-substrate digestion is among others, a function of the mixing ratio which is in turn dependent on the
compositional characteristics of the bioreactor feeds. To improve biogas yield by co-substrate digestion, it is therefore imperative to determine the compositional characteristics of the individual substrates and the ratio at which they must be blended. Optimum performance in anaerobic digestion requires suitable conditions such as mixing, substrate, C/N ratio, pH, temperature, Hydraulic retention time (HRT), and Organic loading rate, have to be established to keep the microbial population in balance [41]. Co-digestion dilutes the inhibitory substances in substrates, balances the micro and macronutrients, increases the organic loading with concomitant higher biogas yield per unit of digester volume; lastly, diversify and synergizes the microbial communities which play a pivotal role in the methanogenesis [42].

![Graph showing anaerobic digestion pattern and daily biogas production from mixtures of PD/CD](image)

**Fig. 2. Anaerobic digestion pattern and daily biogas production from mixtures of PD/CD**

<table>
<thead>
<tr>
<th>Parameters (%</th>
<th>PD</th>
<th>CD</th>
<th>PM</th>
</tr>
</thead>
<tbody>
<tr>
<td>Moisture content (MC)</td>
<td>12.36</td>
<td>9.55</td>
<td>12.38</td>
</tr>
<tr>
<td>Ash content</td>
<td>18.80</td>
<td>30.68</td>
<td>20.14</td>
</tr>
<tr>
<td>Fibre</td>
<td>21.09</td>
<td>30.30</td>
<td>30.99</td>
</tr>
<tr>
<td>Nitrogen</td>
<td>5.60</td>
<td>1.45</td>
<td>3.45</td>
</tr>
<tr>
<td>Crude Protein</td>
<td>35.00</td>
<td>9.06</td>
<td>21.58</td>
</tr>
<tr>
<td>C/N ratio</td>
<td>10.00</td>
<td>37.00</td>
<td>15.00</td>
</tr>
<tr>
<td>Fat content</td>
<td>4.46</td>
<td>2.89</td>
<td>3.92</td>
</tr>
<tr>
<td>Total Organic Carbon</td>
<td>53.00</td>
<td>54.26</td>
<td>50.00</td>
</tr>
<tr>
<td>Total Solids (TS)</td>
<td>87.64</td>
<td>90.45</td>
<td>87.63</td>
</tr>
<tr>
<td>Volatile Solids (VS)</td>
<td>68.84</td>
<td>59.77</td>
<td>67.62</td>
</tr>
</tbody>
</table>

**Table 2. Proximate composition of the substrates**
Fig. 3. Anaerobic digestion pattern and daily biogas production from mixtures of CD/PM

Fig. 4. Cumulative biogas yield from the treatments and the percentage increase and inhibition
Fig. 5. Simulation of experimental data from of PD/CD ratios with modified Gompertz model.

Fig. 6. Simulation of experimental data from of CD/PM ratios with modified Gompertz model.
Fig. 7. Simulation of experimental data from of CD/PM ratios with bi-logistic function model

Fig. 8. Simulation of experimental data from of PD/CD ratios with bi-logistic function model
Table 3. The kinetic constants estimated using modified gompertz model

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Ym (dm$^3$)</th>
<th>U (dm$^3$)</th>
<th>λ (Day)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PD/CD 1:1</td>
<td>38.38</td>
<td>0.70</td>
<td>9.34</td>
</tr>
<tr>
<td>PD/CD 2:1</td>
<td>112.46</td>
<td>1.12</td>
<td>4.55</td>
</tr>
<tr>
<td>PD/CD 3:1</td>
<td>37.67</td>
<td>0.96</td>
<td>13.31</td>
</tr>
<tr>
<td>PD</td>
<td>41.43</td>
<td>0.65</td>
<td>16.87</td>
</tr>
<tr>
<td>CD/PM 1:1</td>
<td>25.79</td>
<td>0.86</td>
<td>11.73</td>
</tr>
<tr>
<td>CD/PM 2:1</td>
<td>22.56</td>
<td>0.56</td>
<td>10.32</td>
</tr>
<tr>
<td>CD/PM 3:1</td>
<td>23.56</td>
<td>0.75</td>
<td>11.08</td>
</tr>
<tr>
<td>CD</td>
<td>21.67</td>
<td>0.46</td>
<td>8.56</td>
</tr>
</tbody>
</table>

Table 4. Kinetic constants estimated using bi-logistic function model

<table>
<thead>
<tr>
<th>Treatment Ratio</th>
<th>Bi-Logistic model parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$Pb_1$ (dm$^3$)</td>
</tr>
<tr>
<td>PD/CD 1:1</td>
<td>15.88</td>
</tr>
<tr>
<td>PD/CD 2:1</td>
<td>8.91</td>
</tr>
<tr>
<td>PD/CD 3:1</td>
<td>16.63</td>
</tr>
<tr>
<td>PD Alone</td>
<td>11.96</td>
</tr>
<tr>
<td>CD/PM 1:1</td>
<td>10.46</td>
</tr>
<tr>
<td>CD/PM 2:1</td>
<td>9.11</td>
</tr>
<tr>
<td>CD/PM 3:1</td>
<td>10.98</td>
</tr>
<tr>
<td>CD Alone</td>
<td>7.95</td>
</tr>
</tbody>
</table>

Table 5. Goodness of fit for the experiments

<table>
<thead>
<tr>
<th>Experiment</th>
<th>Modified Gompertz Model</th>
<th>Bi-logistic Function Model</th>
</tr>
</thead>
<tbody>
<tr>
<td>PD/CD 1:1</td>
<td>0.9797</td>
<td>0.9985</td>
</tr>
<tr>
<td>PD/CD 2:1</td>
<td>0.9968</td>
<td>0.9967</td>
</tr>
<tr>
<td>PD/CD 3:1</td>
<td>0.9877</td>
<td>0.9970</td>
</tr>
<tr>
<td>PD</td>
<td>0.9913</td>
<td>0.9978</td>
</tr>
<tr>
<td>CD/PM 1:1</td>
<td>0.9880</td>
<td>0.9942</td>
</tr>
<tr>
<td>CD/PM 2:1</td>
<td>0.9889</td>
<td>0.9981</td>
</tr>
<tr>
<td>CD/PM 3:1</td>
<td>0.9931</td>
<td>0.9977</td>
</tr>
<tr>
<td>CD</td>
<td>0.9901</td>
<td>0.9920</td>
</tr>
</tbody>
</table>

The characteristics of the three animal manures used in this study as shown in the results, especially the C/N ratio, TS, and VS content suggest their suitability as feedstock in anaerobic digestion and biogas production. Cow dung, being excreta from a ruminant animal is known to contain the autochthonous microbial flora that aids in faster biogas production. It has also been severally reported that cow dung is a very good starter for poor biogas-producing feedstocks [43]. The high VS and nitrogen content, coupled with the low C/N ratio of the poultry manure makes it a suitable substrate for co-digestion with other substrates with high carbon but low nitrogen content [1]. Though the cumulative biogas yield from PD alone is relatively high compared to CD alone, the low C/N ratio of PD underscores the need to improve the compositional characteristics of PD through co-digestion with a nitrogen-rich substrate to further enhance biogas production.

As indicated by the plots of anaerobic digestion (AD) and daily biogas production from the different treatments, microbial activities followed a similar trend in all the digesters. The initial lag phase was above 10 days in some treatments, followed by an active period of biogas production which subsequently declined and finally stopped. Biogas was low at the beginning and the end of the experiment; this implies that the biogas produced in batch conditions corresponds to the specific growth rate of methanogenic bacteria [20]. To be fully effective, microbial populations in
bioreactors and their adaptation to the new environmental conditions typically take a significant period to establish themselves [44].

At the initial stage of AD, the aerobic bacteria use up the available O₂ trapped in the bioreactor to breakdown complex organic compounds into simpler forms, releasing CO₂. As the amount of O₂ available in the bioreactor is being used up, there is a corresponding decrease in the amount of CO₂ produced until all the oxygen is completely exhausted. At this point, the aerobic bacteria are succeeded by the anaerobic and facultative anaerobes, and this halts their activities and methanogenic activities take over the stage. The methane forming bacteria have a very slow growth rate, explaining the gradual rise in gas production after the initial low yield [19]. It could also be explained that when the bioreactors were initially charged, acid forming-bacteria quickly produced acid which results in a decline in pH below neutral, which decreases the growth of methanogenic bacteria and consequently, methanogenesis [45].

In the course of the AD of the mixtures of the feedstocks, biogas production started after a period of relative inactivity (lag phase). The initial rise in gas production was followed by a period of steady-state in gas production, and subsequently, an acceleration in gas yield before it slowly declined. The result of this pattern was multiple peaks in biogas production as indicated in the plots. This phenomenon suggested a biphasic or diauxic-like behavior in the microbial growth and utilization of the complex mixture of substrates in the bioreactors, as was deduced from the successive appearance of multiple peaks in the biogas production profiles. The phenomenon whereby a microbial population, when presented with two or more carbon and energy sources, exhibits biphasic exponential growth intermitted by a period of lag-phase of minimal growth is knowns as diauxic or biphasic growth [33]. The diauxic growth pattern and biogas production profile demonstrate the complex and multi-component nature of the organic wastes in the bioreactors. The presence of compounds such as lipids, carbohydrates, and proteins, which are precursors of intermediate inhibitory compounds, such as long-chain fatty acids and ammonia could be responsible for the observed diauxic growth pattern and bi-phasic biogas production [46].

Diauxic growth pattern for several complex organic substrates such as substrates rich in fat, protein, or lignocellulose has been reported [47], and is mostly ascribed to the fact that microbial populations are exposed to two or more substrates which are preferentially metabolized at different rates, resulting in a two-phase reaction. Though, biphasic or diauxic-like biogas production kinetics may also reveal some level of inhibitory effects on the biochemical steps of the biogas production process. A similar pattern of diauxic growth in the utilization of organic substrates observed in this study has been reported by [48], in which a non-linear model of hydrogen production by Caldicellulosiruptor saccharolyticus for diauxic-like consumption of lignocellulosic sugar mixtures was investigated.

The biphasic or diauxic growth pattern and biogas production profile prompted the application of modified Gompertz model and bi-logistic function model in the evaluation of the AD process, to ascertain the possibility of capturing the observed curves. And as can be seen in Figs. 4-7, the anaerobic digestion process was suitably described by the modified Gompertz model, however, the bi-logistic function model showed a better performance in the simulation of the generated experimental data, because it was able to capture the curves and therefore, most suitably described the AD process with higher correlation coefficient R² than modified Gompertz model (Table 5). The high correlation coefficients > 0.99 indicate a very strong relationship between experimental data and model parameters. Two sets of kinetic parameters, (Pb₁; Rm₁; λ₁ and Pb₂; Rm₂; λ₂) were generated using the bi-logistic model. The Pb₂ (the maximum biogas potential of the substrate) was significantly higher than Pb₁, whereas the Rm₁ (maximum biogas production rate) is lower than the Rm₂. However, the λ₁ (the predicted lag phase) is much higher than the λ₂. This revealed that the different substrate components biodegraded at different rates (Rm), with different biogas potentials (Pb). The phenomenon of diauxic growth in the anaerobic digestion of complex organic substrates could be captured quantitatively in a kinetic model using bi-logistic function model.

5. CONCLUSION

Anaerobic digestion and biogas production technology have proven to be the future of sustainable and eco-friendly agricultural and organic matter-rich industrial waste management pathway. This study showed that co-digestion of PD/CD and CD/PM increased biogas production
at the ratios of 1:1 and 3:1, whereas 2:1 exhibited inhibitory effects. Improvement of biogas production by co-substrate digestion is, among other factors, a function of the mixing ratio which in turn depends on the physicochemical characteristics of the bioreactor feeds. To enhance biogas yield by co-substrate digestion, it is therefore imperative to determine the compositional characteristics of the individual substrates and the ratio at which they must be mixed.

The two models showed good performance in the simulation of the AD process, with high correlation coefficients, an indication of a very strong relationship between experimental data and model parameters. However, the bi-logistic function model showed a better performance in the simulation of the generated experimental data, as it was able to capture the curves in the plots, with a higher correlation coefficient $R^2$ than the modified Gompertz model. One of the findings in this work has shown that the phenomenon of diauxic growth in the anaerobic digestion of complex organic substrates could be captured quantitatively in a kinetic model using the bi-logistic function model. While we recommend further studies in AD using the bi-logistic function model, the preliminary results in this research could be valuable in planning for anaerobic digestion of animal manure for biogas production in large scale.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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